

Seasonal variations on total fatty acid composition of fillets of zander (*Sander lucioperca*) in Beysehir Lake (Turkey)

G.O. Guler^a, A. Aktumsek^{b,*}, O.B. Cital^c, A. Arslan^b, E. Torlak^d

^a Department of Biological Education, Education Faculty, Selcuk University, Konya 42090, Turkey

^b Department of Biology, Science and Arts Faculty, Selcuk University, Konya 42079, Turkey

^c Department of Animal Nutrition, Veterinary Faculty, Selcuk University, Konya 42079, Turkey

^d Provincial Control Laboratory, Konya, Turkey

Received 8 December 2005; received in revised form 21 July 2006; accepted 13 October 2006

Abstract

Seasonal variations on total fatty acid compositions of zander, *Sander lucioperca* in Beysehir Lake, were determined by using GC. Polyunsaturated fatty acids (PUFA) were found to be higher than saturated (SFA) and monounsaturated fatty acids (MUFA) in all seasons. Palmitic acid was the major SFA (57.0–64.0% of total SFA) in all seasons. Oleic acid was identified as the major MUFA (45.0–58.0% of total MUFAs). Docosahexaenoic acid (DHA), linoleic acid (LA), eicosapentaenoic acid (EPA), and arachidonic acid (AA) were the most abundant PUFA. Relating to the total fatty acid composition of zander, the percentages of DHA, LA, AA, and EPA ranged between 17.1–23.3%, 5.40–15.4%, 6.72–9.94% and 4.22–5.93% of total lipid, respectively. The percentages of total ω 3 fatty acid were higher than those of total ω 6 fatty acid in the fatty acid composition of zander with ω 3/ ω 6 ratios of 1.49, 1.45, 1.22, 0.72 in spring, autumn, winter, and summer, respectively. It was shown that the fatty acid composition and ω 3/ ω 6 fatty acids ratio in the muscle of zander were significantly influenced by spawning and season.

© 2006 Elsevier Ltd. All rights reserved.

Keywords: Zander; *Sander lucioperca*; Fatty acid composition; Seasonal variations; Beysehir Lake

1. Introduction

Since it was noted that the reduced cardiovascular mortality of Eskimos was related to fish consumption, fish oil has become subject of thorough investigation (Lancet, 1983). During recent years, fish lipids have been focused on as being beneficial for human health (Anon, 1992). The potential health benefits related to fish consumption are due to the presence of proteins, unsaturated essential fatty acids, minerals, and vitamins. Additional health benefits from the consumption of fish or fish oil may be related to polyunsaturated fatty acids (PUFAs), especially ω 3 PUFAs (Sidhu, 2003). The composition of fish oils is dominated by two members of the

ω 3 PUFA family: 20:5 ω 3 (eicosapentaenoic acid, known as EPA) and 22:6 ω 3 (docosahexaenoic acid, known as DHA). Long chain ω 3 PUFAs cannot be readily synthesised by human bodies and mostly are obtained through the diet (Alasalvar, Taylor, Zubcov, Shahidi, & Alexis, 2002). Thus, PUFA, especially the longer-chain ω 3 and ω 6 PUFA, have been considered essential fatty acids and have been shown to have curative and preventive effects on cardiovascular diseases, neurodevelopment in infants, cancers and fat glycemic control (Conner, 1997; Kinsella, Lokesh, & Stone, 1990). The ω 3 fatty acids are always present in fish flesh even in lean fish (Ackman, 2002). Results of clinical and epidemiological research suggest that EPA and DHA, found mainly in fish and seafoods have extremely beneficial properties for the prevention of human coronary artery disease (Leaf & Weber, 1988). The evidence of fish oil's heart protective benefits is strong enough that the American

* Corresponding author.

E-mail address: aktumsek@selcuk.edu.tr (A. Aktumsek).

heart association (AHA) now urges everyone to eat at least two servings of fish a week, along with other foods rich in alpha linolenic acid. In the fall of 2002 the AHA advised people who already have heart disease to consume about 1 gram a day of the active ingredients in fish oil—EPA and DHA. Additionally, patients needing to lower elevated triglyceride levels are recommended to consume 2–4 g EPA + DHA daily under the guidance of a physician (Kris-Etherton, Harris, & Appel, 2002). Lemaitre et al. (2003) reported that higher blood EPA plus DHA levels was associated with a 70% lower risk of fatal ischemic heart disease in older adults.

The amount of longer-chain ω 3 PUFAs differs between species and can be influenced by a number of factors. The fatty acid composition of fish tissue can be affected by diet, size, age, reproductive cycle, salinity, temperature, season and geographical location (Bandarra, Batista, Nunes, Empis, & Christie, 1997; Henderson & Tocher, 1987; Leger, Bergot, Lukueta, Flanzky, & Meurot, 1977; Luzia, Sampaio, Castellucci, & Torres, 2003; Shirai, Suzuki, Tokairin, & Wada, 2001, 2002).

Herbivorous fish, which feed on algae containing large amounts of C18 PUFAs but lesser amounts of C20 and C22 PUFAs (Henderson & Tocher, 1987) are found to be rich in ω 6 PUFAs and shorter-chain ω 3 PUFAs (Brown, Roberts, & Truswell, 1989). Carnivores, due to their consumption of other fish in which chain elongation and desaturation is completed, were rich in longer-chain ω 3 PUFAs but lower in α -linolenic acid. Omnivorous fish have a higher proportion of C18 linolenic acid but a lower proportion of longer-chain ω 3 PUFAs (Brown et al., 1989), thus reflecting their diet.

Cold water species have higher levels of longer-chain highly unsaturated ω 3 fatty acids in their diets than warm-water species, which have higher levels of ω 6 and ω 9 fatty acids (Lovell, 1991). Generally a decrease in temperature results in an increase in the degree of unsaturation (Henderson & Tocher, 1987). A possible explanation is that at low temperatures a higher degree of fatty acid unsaturation is needed to maintain flexibility and permeability in membrane phospholipids (Lovell, 1991).

The most important change in total lipid and fatty acid composition of fish is observed during the period of reproduction. In this period, the storage lipids as well as other nutritional compounds such as proteins, vitamins, and minerals in muscle, liver, and visceral organs are mobilized to the gonads in order to ensure maturation (Agren, Muje, Hanninen, Herranen, & Penttila, 1987; Cejas et al., 2003). Therefore, the nutritional quality of muscle may decrease during gonadal maturation (Uysal & Aksoylar, 2005).

The aim of this study was to determine the seasonal variations in total fatty acid composition and ω 3/ ω 6 fatty acids ratio of muscle of zander, *Sander lucioperca*, which is of great commercial importance in Beysehir Lake, the largest freshwater lake in Turkey. Fishing is mostly between July and March.

2. Materials and methods

2.1. Materials

S. lucioperca (zander), used in this study, were obtained seasonally from Beysehir Lake which is the largest freshwater lake (651 km²) in Turkey. Beysehir Lake is located in central Anatolia and 90 km from the city of Konya (37° 45' North–31°30' East). Its average depth is 7–8 m, length about 45 km and width is 14–26 km. It is tectonic/karstic in origin. The water of the lake is used for irrigation and drinking purposes. Zander is one of the most abundant fish in all seasons in this lake and has a great commercial importance for central anatolian peoples.

Zander is a carnivorous fish, and it generally feeds on other fish species rich in fatty acids (Ekmekçi, Erkakan, & Bayrak, 1991). Zander is an important freshwater fish species distributed especially in Eastern and Central Europe, Western Asia, as well as in many other countries in the world. In Turkey, especially in Central Anatolia, zander is a commercial fish species. Zander was implanted into Beysehir Lake between 1978 and 1980 (Erdem, Sarhan, & Erdem, 1985), and no reports have yet been published about the effects of seasonal variations on the fatty acid composition of this important species in this location.

The seasons chosen for analysis were summer, winter, spring, and autumn. The samples were collected in middle month of each season during 2003. All representative fishes ($n = 3$ at each determination) used in the experiments were of almost the same size and age. After being caught, they were transported on ice to the laboratories, filleted, and then frozen. At the beginning of each analysis, the samples were allowed to equilibrate to room temperature, ground, and homogenized in chloroform/methanol mixture (2/1 v/v).

2.2. Fatty acid analysis

Samples of fillets were extracted by the Folch, Lees, and Sloane Stanley (1957) method were transesterified with BF₃-methanol (Moss, Lambert, & Merwin, 1974).

Fatty acid methyl esters (FAMES) were analyzed on a HP (Hewlett Packard) Agilent 6890N model gas chromatograph (GC), equipped with a flame ionization detector (FID) and fitted with a DB-23 capillary column (60 m, 0.25 mm i.d. and 0.25 μ m). Injector and detector temperatures were 270 and 280 °C, respectively. Column temperature program was 190 °C for 35 min then increasing at 30 °C/min up to 220 °C where it was maintained for 5 min. Carrier gas was helium (2 ml/min) and split ratio was 30:1.

Identification of normal fatty acids was carried out by comparing sample FAME peak relative retention times with those obtained for Alltech standards. Results were expressed as FID response area relative percentages.

2.3. Statistical analysis

Each reported result is the average value of three GC analyses. The results are offered as means \pm SD. The results were submitted to analysis of variance (ANOVA), at 0.05 significance level, using SPSS, 10.0. The mean values were compared by Tukey's test.

3. Results and discussion

Table 1 shows the lipid content of the fillets of zander investigated from the Beysehir Lake of Turkey. The lipid content ranged from 0.58% to 1.26% in fillets of zander. Similarly, Jankowska, Zakes, Zmijewski, and Szczepkowski (2003) found that contents of fat in the zander muscle tissue was 0.96% and Uysal and Aksoylar (2005) found that total lipid of muscle of zander was between 0.39% and 0.77%.

Seasonal variations on total fatty acid composition of zander are presented in Table 2. We identified and evaluated 35 fatty acids in muscle lipids of zander. The major fatty acids in the zander in all seasons were 22:6 ω 3 (DHA), 16:0, 18:1 ω 9, 20:4 ω 6 (AA), 18:2 ω 6, 16:1 ω 7, 20:5 ω 3 (EPA), and 18:0, respectively.

Palmitic acid was the major SFA, contributing approximately 57.0–64.0% to the total SFA content of the lipids for zander. Similar results for zander (Çelik, Diler, & Küçükgülmez, 2005; Jankowska et al., 2003), and other freshwater fish (Haliloğlu, Aras, & Yetim, 2002; Rahman, Huah, Hassan, & Daud, 1995) have also been reported. Ackman, Eaton, and Linne (1975) pointed out that palmitic acid was a key metabolite in fish and its level was not influenced by diet.

Oleic acid was identified as the major MUFA in the fish (45.0–58.0% of total MUFAs). Oleic acid in muscle tissue of zander was found to be 9.10%, 8.30%, 10.5%, and 11.9% in spring, summer, autumn, and winter, respectively. Similarly, Haliloğlu, Bayır, Sirkecioğlu, Aras, and Atamanalp (2004) found that C18:1 ω 9 was the major MUFA in muscle in tissue of rainbow trout (*Oncorhynchus mykiss*) living in freshwater. The high levels of oleic, palmitoleic, and arachidonic acids had been reported as a characteristic property of freshwater fish oils (Andrade, Rubira, Matsushia, & Souza, 1995; Osman, Suriah, & Law, 2001).

PUFAs were found to be a significant constituent 50.2–57.0% of muscle lipid of zander, according to the season. DHA (C22:6 ω 3), LA (C18:2 ω 6), AA (C20:4 ω 6) and EPA

(C20:5 ω 3) were the predominant PUFAs. The present data showed that DHA (C22:6) was the predominant fatty acid in muscle lipids of zander, and its level showed the most variation during seasons, accounting for 17.1–23.3% of total fatty acids and it was determined at 23.3%, 17.1%, 22.6% and 18.7% in spring, summer, autumn and winter, respectively. Similarly, Jankowska et al. (2003) found that DHA was the predominant fatty acid (24.5%) in muscle lipid tissue of wild zander and in this study, cultured zander fed artificial feed has more total fat (2.87%) than wild zander (0.96%). Sargent (1996) reported that ω 3 PUFA, principally DHA, has a role in maintaining the structure and functional integrity of fish cells. In addition, DHA has a specific and important role in neural cell membranes, i.e. the brain and eyes. Moreover, it is considered a desirable property in fish for human nutrition and health.

In the richer feeding period, zander partially preferred to accumulate PUFA rather than SFA and MUFA. In this study, it was found that total PUFA in the fatty acid composition of zander muscle in winter was low where we had hoped that the percentage of total PUFA would be high. The observed decrease in PUFA is likely due to their utilization for gonad maturation. The gonads were not included in the analysis of the material extracted. This conclusion is also supported by the fact that the level of PUFA was lowest both before and just after reproduction (Agren et al., 1987; Cejas et al., 2003). In the same way, Karakaya and Kılıç (1995) reported that zander in Beysehir Lake probably draw near to reproductive period in February.

In the present study, the percentages (in total lipid) of EPA and DHA which have a vital role in human nutrition were between 4.22–5.93% and 17.1–23.3%, respectively, according to the seasons. Thus, among the ω 3 series, the zander are good sources of EPA and DHA in all seasons.

The main characteristic difference in freshwater fish is the higher levels of C-16 and C-18 acids and the lower levels of C-20 and C-22 acids when compared to marine fish, and these differences are mainly due to the dietary fat (Ackman, 1967); however freshwater fish contain relatively large amounts of EPA and DHA (Wang, Miller, Perren, & Addis, 1990). It should be pointed out that the PUFA contents in the fish studied in this work were higher than those reported by Uysal and Aksoylar (2005) from zander inhabiting Eğirdir Lake, which is the second largest freshwater lake in Turkey. The discrepancy was primarily caused by a higher DHA content. The results obtained in this study show shorter (C:18) chain ω 3 acids in the food to be elongated and desaturated in the zander body, whereby longer-chain PUFAs, mainly DHA, are formed. The results demonstrate that zander is highly capable of transforming native forms of 18 ω 3 into long-chain acids, as a result of which the muscle has a high DHA content. Similar results were obtained by Xu, Fontaine, Melard, and Kestemont (2001) who analysed dietary effects on fatty acid composition in muscles and liver of *Perca fluviatilis*; they found

Table 1
Lipid values determined in spring, summer, autumn and winter for the zander species investigated

Seasons	Lipids (% wet weight basis)
Spring	0.62
Summer	0.8
Autumn	0.58
Winter	1.26

Table 2
Seasonal variations on total fatty acid composition of zander (*Sander lucioperca*) in Beysehir Lake*

Fatty acids	Spring	Summer	Autumn	Winter
C8:0	–	0.06 ± 0.12 ^{a**}	–	–
C10:0	–	–	–	0.01 ± 0.18
C12:0	0.03 ± 0.2 ^a	0.14 ± 0.15 ^a	0.12 ± 0.11 ^a	0.07 ± 0.11 ^a
C13:0	–	0.01 ± 0.52 ^a	0.02 ± 0.2 ^a	0.07 ± 0.1 ^a
C14:0 ^{***}	0.65 ± 0.19 ^a	0.67 ± 0.17 ^a	0.89 ± 0.25 ^a	1.45 ± 0.67 ^b
C15:0	0.20 ± 0.2 ^a	0.18 ± 0.17 ^a	0.25 ± 0.15 ^b	0.30 ± 0.2 ^b
C16:0	14.4 ± 0.89 ^a	17.9 ± 2.85 ^b	16.5 ± 0.87 ^b	14.2 ± 1.63 ^a
C17:0	0.49 ± 0.12 ^a	0.60 ± 0.11 ^a	0.63 ± 0.11 ^a	0.72 ± 0.57 ^a
C18:0	4.76 ± 0.66 ^a	6.21 ± 0.36 ^b	5.29 ± 0.78 ^a	4.08 ± 0.21 ^c
C19:0	0.06 ± 0.2 ^a	0.01 ± 0.2 ^a	0.08 ± 0.2 ^a	0.11 ± 0.15 ^a
C20:0	0.03 ± 0.2 ^a	0.15 ± 0.14 ^a	0.10 ± 0.11 ^a	–
C21:0	0.03 ± 0.2 ^a	–	0.12 ± 0.18 ^a	–
C24:0	3.82 ± 0.63 ^a	2.02 ± 0.22 ^b	3.28 ± 0.75 ^a	3.86 ± 0.45 ^a
∑SFA	24.4	27.9	27.3	24.8
C14:1 ω5	0.11 ± 0.2 ^a	0.17 ± 0.15 ^a	0.31 ± 0.15 ^a	0.42 ± 0.47 ^a
C15:1 ω6	0.24 ± 0.2 ^a	0.47 ± 0.66 ^a	0.21 ± 0.16 ^a	0.17 ± 0.18 ^a
C16:1 ω7	6.75 ± 1.51 ^a	2.31 ± 1.01 ^b	3.40 ± 0.64 ^b	7.18 ± 1.58 ^a
C17:1 ω8	0.85 ± 0.2 ^a	0.71 ± 0.41 ^a	0.82 ± 0.17 ^a	0.52 ± 0.32 ^a
C18:1 ω9	9.10 ± 2.4 ^a	8.30 ± 0.42 ^a	10.53 ± 1.13 ^b	11.9 ± 0.43 ^c
C18:1 ω7	2.90 ± 0.5 ^a	2.07 ± 0.45 ^b	2.81 ± 0.27 ^a	3.43 ± 0.18 ^c
C20:1 ω9	0.27 ± 0.2 ^a	0.08 ± 0.2 ^a	0.15 ± 0.17 ^a	–
C24:1 ω9	–	0.21 ± 0.29 ^a	0.08 ± 0.11 ^a	–
∑MUFA	20.2	14.3	18.3	23.6
C16:2 ω4	0.92 ± 0.58 ^a	0.83 ± 0.41 ^a	0.70 ± 0.17 ^a	0.70 ± 0.44 ^a
C18:2 ω6	5.40 ± 0.74 ^a	15.4 ± 2.82 ^b	7.73 ± 3.13 ^a	7.54 ± 1.92 ^a
C18:3 ω6	0.88 ± 0.24 ^a	1.02 ± 0.64 ^a	1.14 ± 0.2 ^a	2.11 ± 0.89 ^b
C18:3 ω3	0.32 ± 0.2 ^a	0.11 ± 0.16 ^a	0.24 ± 0.4 ^a	0.62 ± 0.48 ^b
C20:2 ω6	–	0.18 ± 0.15 ^a	0.06 ± 0.2 ^b	–
C20:3 ω6	–	–	0.10 ± 0.15 ^a	–
C20:3 ω3	–	0.08 ± 0.15 ^a	0.11 ± 0.16 ^a	0.06 ± 0.17 ^a
C20:4 ω6	9.94 ± 1.29 ^a	6.72 ± 0.36 ^b	9.67 ± 1.85 ^a	8.68 ± 1.38 ^a
C20:5 ω3	5.93 ± 0.46 ^a	4.22 ± 0.84 ^b	5.67 ± 0.19 ^a	5.54 ± 0.31 ^a
C22:3 ω3	–	0.30 ± 0.44 ^a	0.24 ± 0.36 ^a	–
C22:4 ω6	1.64 ± 0.14 ^a	0.93 ± 0.12 ^b	1.52 ± 0.68 ^a	1.80 ± 0.40 ^a
C22:5 ω3	2.53 ± 0.24 ^a	1.81 ± 0.36 ^b	2.31 ± 0.41 ^a	2.42 ± 0.25 ^a
C22:5 ω6	3.36 ± 1.28 ^a	8.28 ± 3.95 ^b	0.99 ± 0.98 ^a	2.04 ± 2.19 ^a
C22:6 ω3	23.3 ± 3.4 ^a	17.1 ± 3.63 ^b	22.6 ± 1.28 ^a	18.7 ± 3.94 ^b
∑PUFA	54.2	57.0	53.1	50.2
Unknown	1.19	0.77	1.34	1.36
ω3	32.0	23.6	31.2	27.4
ω6	21.5	33.0	21.4	22.3
ω3/ω6	1.49	0.72	1.45	1.22

* Average of three lots analysed.

** Values reported are means ± S.D.

*** abc Values for each sample with different superscript letters in the same fraction are significantly different at $P < 0.05$.

high DHA contents. Similarly, Jankowska et al. (2003) found higher DHA concentrations in zander and they found that the zander meat content of DHA, a fatty acid originating in the fish, was high and independent of the food DHA contents; this shows a potential of (18ω3) PUFA to be transformed into more unsaturated long-chain PUFA.

There are close relationships between the fish lipid composition and the diets of fish (McKenzie et al., 2000). Zander is a carnivorous fish and carnivores due to their consumption of other fish in which chain elongation and desaturation is completed were rich in long chain ω3 PUFAs, but lower in

α-linolenic acid. In our study, PUFA levels of zander in Beysehir Lake were found to be high (50.2–57.0%), and α-linolenic acid level was found to be low (0.11–0.62%). However the γ-linolenic acid (C18:3n–6), presumably an intermediate between linoleic (C18:2n–6) and arachidonic acid (C20:4n–6), is relatively high.

The ω3/ω6 ratio is a good index for comparing relative nutritional value of fish oils (Piggott & Tucker, 1990). It is important for human health to increase the consumption of fish or fish products, which are rich in PUFAs of the ω3 family and poorer in PUFAs of the ω6 family (Sargent, 1997). The present data show that the ω3/ω6 ratio was

1.49 in spring, 1.45 in autumn, 1.22 in winter and that the lowest value 0.72 was in summer. A high level of $\omega 6$ fatty acids lowered the $\omega 3/\omega 6$ ratio in summer.

This study has shown that the zander is suitable item in the human diet during the fishing period in the Beyşehir Lake of Turkey when the levels of EPA, DHA and $\omega 3/\omega 6$ ratio are considered. This condition can be regarded as an explanation for the fact that the zander in Beyşehir Lake are richer in $\omega 3$ fatty acids, taking into consideration with the fatty acid profile of the fish. As a consequence, when human health is taken into account, the zander from Beyşehir Lake appears to be quite nutritious in terms of fatty acid composition and ratio.

Acknowledgements

This study was financed by Selcuk University Scientific Research Foundation (BAP) under Project FBE 2003/189. The authors acknowledge their support of this Project.

References

- Ackman, R. G. (1967). Characteristics of the fatty acid composition and biochemistry of some fresh-water fish oils and lipids in comparison with marine oils and lipids. *Comparative Biochemistry and Physiology*, *22*, 907–922.
- Ackman, R. G. (2002). Freshwater fish lipids- an overlooked source of beneficial long-chain $n-3$ fatty acids. *European Journal of Lipid Science and Technology*, *104*, 253–254.
- Ackman, R. G., Eaton, C. A., & Linne, B. A. (1975). Differentiation of freshwater characteristics of fatty acids in marine specimens of the atlantic sturgeon (*Acipenser oxyrinchus*). *Fishery Bulletin*, *73*, 838–845.
- Agren, J., Muje, P., Hanninen, O., Herranen, J., & Penttila, I. (1987). Seasonal variations of lipid fatty acids of Boreal freshwater fish species. *Comparative Biochemistry and Physiology*, *88*, 905–909.
- Alasalvar, C., Taylor, K. D. A., Zubcov, E., Shahidi, F., & Alexis, M. (2002). Differentiation of cultured sea bass (*Dicentrarchus labrax*): Total lipid content, fatty acid and trace mineral composition. *Food Chemistry*, *79*(2), 145–150.
- Andrade, A. D., Rubira, A. F., Matsushia, M., & Souza, N. E. (1995). Omega3 fatty acids in freshwater fish from South Brazil. *Journal of the American Oil Chemists Society*, *72*(10), 1207–1210.
- Anon. (1992). *Unsaturated fatty acids. Nutritional and physiological significance. British nutrition foundation report* (pp. 156–157). The Report of the British Nutrition Foundation's Task Force. Chapman & Hall, London.
- Bandarra, N. M., Batista, I., Nunes, M. L., Empis, J. M., & Christie, W. W. (1997). Seasonal changes in lipid composition of Sardine (*Sardina pilchardus*). *Journal of Food Science*, *62*(1), 40–42.
- Brown, A. J., Roberts, D. C. K., & Truswell, A. S. (1989). Fatty acid composition of Australian marine finfish: A review. *Food Australia*, *41*(3), 655–666.
- Çejas, J. R., Almansa, E., Villamandos, J. E., Badia, P., Bolanos, A., & Lorenzo, A. (2003). Lipid and fatty acid composition of ovaries from wild fish and ovaries and eggs from captive fish of white sea bream (*Diplodus sargus*). *Aquaculture*, *216*(1–4), 299–313.
- Çelik, M., Diler, A., & Küçükgülmez, A. (2005). A comparison of the proximate compositions and fatty acid profiles of zander (*Sander lucioperca*) from two different regions and climatic conditions. *Food Chemistry*, *92*, 637–641.
- Conner, W. E. (1997). The beneficial effects of omega-3 fatty acids: cardiovascular disease and neurodevelopment. *Current Opinion in Lipidology*, *8*, 1–3.
- Ekmekçi, F.G., Erkakan, F., & Bayrak, M. (1991). Eğirdir Gölü stok tespiti 1990 yılı kesin raporu. *TÜBİTAK DEĞAÇ97/G*, Ankara, 115 s.
- Erdem, Ü., Sarihan, E., & Erdem, C. (1985). Beyşehir Gölü Sudak (*Stizostedion lucioperca* L. 1758) populasyonlarının meristik özellikleri ile gelişme, boy-uzunluk ilişkisi ve kondüsyon üzerine bir araştırma. *C.Ü Fen Bil. Enst. Derg.*, *3*, 237–252.
- Folch, J., Lees, M., & Sloane Stanley, G. H. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, *226*, 497–509.
- Haliloğlu, H. I., Aras, N. M., & Yetim, H. (2002). Comparison of muscle fatty acids of three trout species (*Salvelinus alpinus*, *Salmo trutta fario*, *Oncorhynchus mykiss*) raised under the same conditions. *Turkish Journal of Veterinary and Animal Sciences*, *26*(5), 1097–1102.
- Haliloğlu, H. I., Bayır, A., Sirkecioğlu, A. N., Aras, N. M., & Atamanalp, M. (2004). Comparison of fatty acid composition in some tissues of rainbow trout (*Oncorhynchus mykiss*) living in seawater and freshwater. *Food Chemistry*, *86*, 55–59.
- Henderson, R. J., & Tocher, D. R. (1987). The lipid composition and biochemistry of freshwater fish. *Progress in Lipid Research*, *26*(4), 281–347.
- Jankowska, B., Zakes, Z., Zmijewski, T., & Szczepkowski, M. (2003). Fatty acid profile and meat utility of wild and cultured zander, *Sander lucioperca* (L.). *Electronic Journal Of Polish Agricultural Universities Fisheries*, *6*(1).
- Karakaya, M., & Kılıç, A. (1995). Beyşehir Gölünden 1994–1995 avlanma periyodunda yakalanan levrek balıklarının bazı özelliklerinde meydana gelen değişimin tesbiti üzerine bir araştırma. *S.Ü. Ziraat Fakültesi Dergisi*, *8*(10), 50–60.
- Kinsella, J. E., Lokesh, B., & Stone, R. A. (1990). Dietary $n-3$ polyunsaturated fatty acids and amelioration of cardiovascular disease: possible mechanisms. *American Journal of Clinical Nutrition*, *52*(1), 1–28.
- Kris-Etherton, P. M., Harris, W. S., & Appel, L. J. (2002). Fish consumption, fish oil, omega-3 fatty acids, and cardiovascular disease. *Circulation*, *106*, 2747–2757.
- Lancet (1983). Eskimo diets and diseases. *Lancet*, *1*, 1139–1141.
- Leaf, A., & Weber, P. C. (1988). Cardiovascular effects of $n-3$ fatty acids. *New England Journal of Medicine*, *318*(9), 549–557.
- Leger, C., Bergot, P., Lukuet, P., Flanzly, J., & Meurot, J. (1977). Specific distribution of fatty acids in the triglycerides of rainbow trout adipose tissue. Influence of temperature. *Lipids*, *12*(7), 538–543.
- Lemaitre, R. N., King, I. B., Mozaffarian, D., Kuller, L. H., Tracy, R. P., & Siscovick, D. S. (2003). $n-3$ polyunsaturated fatty acids, fatal ischemic heart disease, and nonfatal myocardial infarction in older adults: The Cardiovascular Health Study. *American Journal of Clinical Nutrition*, *77*(2), 319–325.
- Lovell, R. T. (1991). Nutrition of aquaculture species. *Journal of Animal Science*, *69*(10), 4193–4200.
- Luzia, L. A., Sampaio, G. R., Castellucci, C. M. N., & Torres, E. A. F. S. (2003). The influence of season on the lipid profiles of five commercially important species of Brazilian fish. *Food Chemistry*, *83*(1), 93–97.
- McKenzie, D. J., Piraccini, G., Piccolella, M., Steffensen, J. F., Bolis, C. L., & Taylor, E. W. (2000). Effects of dietary fatty acid composition on metabolic rate and responses to hypoxia in the European eel (*Anguilla anguilla*). *Fish Physiology and Biochemistry*, *22*(4), 281–296.
- Moss, C. W., Lambert, M. A., & Merwin, W. H. (1974). Comparison of rapid methods for analysis of bacterial fatty acids. *Applied Microbiology*, *28*, 80–85.
- Osman, H., Suriah, A. R., & Law, E. C. (2001). Fatty acid composition and cholesterol content of selected marine fish in Malaysian waters. *Food Chemistry*, *73*(1), 55–60.
- Piggott, G. M., & Tucker, B. W. (1990). *Effects of technology on nutrition*. New York: Marcel Dekker.

- Rahman, S. A., Huah, T. S., Hassan, O., & Daud, N. M. (1995). Fatty acid composition of some Malaysian freshwater fish. *Food Chemistry*, *54*(1), 45–49.
- Sargent, J. R. (1996). Origins and functions of egg lipid. In N. R. Bromage & R. J. Roberts (Eds.), *Broodstock management and egg and larval quality* (pp. 353–372). Oxford: Blackwell.
- Sargent, J. R. (1997). Fish oils and human diet. *British Journal of Nutrition*, *78*(1), S5–S13.
- Shirai, N., Suzuki, H., Tokairin, S., Ehara, H., & Wada, S. (2002). Dietary and seasonal effects on the dorsal meat lipid composition of Japanese (*Silurus asotus*) and Thai catfish (*Clarias macrocephalus* and hybrid *Clarias macrocephalus* and *Clarias galipinus*). *Comparative Biochemistry and Physiology, Part A*, *132*(3), 609–619.
- Shirai, N., Suzuki, H., Tokairin, S., & Wada, S. (2001). Spawning and season affect lipid content and fatty acid composition of ovary and liver in Japanese catfish (*Silurus asotus*). *Comparative Biochemistry and Physiology, Part B*, *129*(1), 185–195.
- Sidhu, K. S. (2003). Health benefits and potential risks related to consumption of fish or fish oil. *Regulatory Toxicology and Pharmacology*, *38*(3), 336–344.
- Uysal, K., & Aksoylar, M. Y. (2005). Seasonal variations in fatty acid composition and the $n-6/n-3$ fatty acid ratio of pikeperch (*Sander lucioperca*) muscle lipids. *Ecology of Food and Nutrition*, *44*, 23–35.
- Wang, Y. J., Miller, L. A., Perren, M., & Addis, P. B. (1990). Omega-3 fatty acids in lake superior fish. *Journal of Food Science*, *55*(1), 71–73.
- Xu, X. L., Fontaine, P., Melard, C., & Kestemont, P. (2001). Effects of dietary fat levels on growth, feed efficiency and biochemical compositions of Eurasian perch *Perca fluviatilis*. *Aquacultural International*, *9*(5), 437–449.